

**Phytochemical and biological overview of genus "Bignonia" (1969-2018)****Basma K. Mahmoud<sup>1</sup>, Ashraf N. E. Hamed<sup>1\*</sup>, Mamdouh N. Samy<sup>1</sup>, Mohamed S. Kamel<sup>1,2</sup>**<sup>1</sup> Department of Pharmacognosy, Faculty of Pharmacy, Minia University, 61519 Minia, Egypt<sup>2</sup> Department of Pharmacognosy, Faculty of Pharmacy, Deraya University, 61111 New Minia, Egypt

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**Abstract**

*Bignonia* L. is the fifth largest genus in the tribe Bignonieae, with 31 lianas species. *Bignonia* plants are widely used in traditional medicine as skin ailments like fungal infections, postpartum haemorrhage, malaria, diabetes and pneumonia. Many literature reported that *Bignonia* species contained different classes of active constituents as phenylethanoids, phenolics, lignans, flavonoids, coumarins and xanthones. Additionally, they demonstrated a wide range of biological activities as cytotoxic, wound healing, antidiabetic, sleep induction, gastroprotection, anti-obesity and insecticidal activities. This review revealed that only 15 species have been reported in the literature. Moreover, *Bignonia* plants need further studies as they are considered a good source for bioactive natural products.

**Key words***Bignonia, Bignoniaceae, Phytochemical, Biological activities, review***1. Introduction**

*Bignoniaceae* is a flowering plants family, comprising of about 110 genera and 650 species, and commonly known as the Trumpet Creeper family [1, 2]. *Bignoniaceae* plants have various bioactive secondary metabolites with diverse pharmacological activities. They are widely used in traditional medicinal systems of a number of countries for the treatment of ailments like cancer, snake bite, skin disorders, gastrointestinal disorders, respiratory tract disorders, hepatic disorders, epilepsy, cholera, pain, urinary problems, malaria, heart problems and sexually transmitted diseases [1]. *Bignonia* L. is the fifth largest genus in the tribe Bignonieae (*Bignoniaceae*), with 31 lianas species distributed from Argentina to USA [3, 4]. Many species are reported to have different classes of bioactive secondary metabolites including; phenylethanoids, phenolics, lignans, flavonoids, coumarins and xanthones [5-7]. *Bignonia* genus has different species with various synonyms. The most popular species are listed in (Table 1) with their synonyms. Most *Bignonia* species are used in folk medicine for treating a wide range of ailments as *B. africana* fruit is used to treat skin ailments like fungal infections, boils, psoriasis, eczema, dysentery, anthelmintic (ringworm and tapeworm), postpartum haemorrhage, malaria, diabetes and pneumonia, while its bark is used to treat venereal diseases and the root is applied to treat ulcer [8, 9]. *B. unguis-cati* is used traditionally to treat a snake bite, dysentery, inflammation, rheumatism, venereal disease and as a quinine substitute for malaria [10-12].

**2. Methodology**

We performed a systematic search on genus "Bignonia" and reported and reported synonyms of different species as

keywords for the search on different databases such as Science direct, DNP, SciFinder, Google Scholar and Scopus. The main search words were "Bignonia" or "biological activity" or "bioactive compounds" or "phytochemical compounds" or "diseases".

**3. Chemical composition**

*Bignonia* genus contained various classes of secondary metabolites, noticed from the isolated groups of compounds including; flavonoids, iridoids and quinones, which were the most abundant among *Bignonia* species. Besides, phenolic acid derivatives, phenylethanoids, lignans, coumarins and sterols. A comprehensive list of the previously isolated compounds from *Bignonia* species is presented in (Table 2 and Figure 1).

**3.1. Flavonoids**

Flavonoids are polyphenolic compounds that possess different health benefits due to their antioxidant potential [13]. Flavonoids are the most abundant in different *Bignonia* plants as *B. brachypoda*, *B. unguis-cati* and *B. callistigoides* [6, 14, 15],....etc.

**3.2. Iridoids**

They are the second abundant class of secondary metabolites isolated from *Bignonia* plants, with different biological activities. In which, verminoside (136), specioside (137) and minecoside (138) were reported in *B. africana* bark and have antiamoebic activity [16].

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**Table 1:** A list of plants belongs to genus "Bignonia" with their synonyms.

Bignonia species	Synonym (s)
<i>B. digitalis</i> Vell.	<i>Zeyheria digitalis</i> (Vell.) Hoehne
<i>B. binata</i> Thunb.	<i>Clytostoma binatum</i> Thunb. Sandwith
<i>B. callistigoides</i> Cham.	<i>Clytostoma callistigoides</i> (Cham.) Baill.
<i>B. capensis</i> Thunb	<i>Tecomaria capensis</i> (Thunb.)
<i>B. adenophylla</i> Wall. & G. Don	<i>Heterophragma adenophyllum</i> (Wall. & G. Don)
<i>B. africana</i> Lam.	<i>Kigelia africana</i> (Lam.) or <i>Kigelia pinnata</i> (Jacq.)
<i>B. unguis-cati</i> L.	<i>Macfadyena unguis-cati</i> (L.) A.H.Gentry
<i>B. hymenaea</i> DC.	<i>Pachyptera hymenaea</i> DC ( <i>Mansao hymenaea</i> )
<i>B. brachypoda</i> DC.	<i>Arrabidaea brachypoda</i> Bureau
<i>B. cuprea</i> Cham.	<i>Arrabidaea chica</i> f. <i>cuprea</i> (Cham.) Sandwith
<i>B. samydoides</i> Cham	<i>Arrabidaea samydoides</i> (Cham.) Sandwith
<i>B. triplinervia</i> Mart. ex DC.	<i>Arrabidaea triplinervia</i> (Mart. ex DC.) Baill.
<i>B. patellifera</i> Schltdl.	<i>Arrabidaea patellifera</i> (Schltdl.) Sandwith
<i>B. crucigera</i> L.	<i>Pithecoctenium crucigerum</i> (L.) A.H.Gentry
<i>B. elegans</i> Vell.	<i>Pseudocalymma elegans</i> (Vell.)
<i>B. stans</i> L.	<i>Tecoma stans</i> (L.) Kunth
<i>B. aesculifolia</i> Kunth	<i>Godmania aesculifolia</i> Standl

**Table 2:** A list of previously reported compounds from genus "Bignonia" (with different synonyms as shown in Table 1).

Classification	No.	Item	Part used	Source	Ref.
<b>I-Phenylethanoids</b>					
	<b>1</b>	Decaffeoyl acteoside	Aerial part	<i>B. unguis-cati</i>	[14]
	<b>2</b>	Verbascoside	Leaf	<i>B. stans</i>	[19]
	<b>3</b>	Isoverbascoside	Stem	<i>B. crucigera</i>	[5]
	<b>4</b>	Forsythoside B	Stem	<i>B. crucigera</i>	[5]
	<b>5</b>	Jionoside D			
	<b>6</b>	Leucosceptoside B			
<b>II-Phenolic compounds</b>					
	<b>7</b>	4-Dihydroxyphenyl)ethyl 2-O-[5-O-(4-hydroxy-3,5-dimethoxybenzoyl)-β-D-apiofuranosyl]-β-D-glucopyranoside	Root	<i>B. brachypoda</i>	[6]
	<b>8</b>	3,4-Dihydroxyphenyl)ethyl 2-O-[5-O-(3,4-dihydroxybenzoyl)-β-D-apiofuranosyl]-β-D-glucopyranoside			
	<b>9</b>	<i>p</i> -Coumaric acid	Stem bark	<i>B. africana</i>	[18]
	<b>10</b>	Caffeic acid			
	<b>11</b>	Ferulic acid	Stem and fruit	<i>B. africana</i>	[20]
	<b>12</b>	Vanillic acid	Stem wood	<i>B. digitalis</i>	[21]
	<b>13</b>	Veratric acid			
	<b>14</b>	Atranorin	Stem bark	<i>B. africana</i>	[22]
	<b>15</b>	Chlorogenic acid	Leaf	<i>B. stans</i>	[19]
	<b>16</b>	Isochlorogenic acid	Leaf	<i>B. unguis-cati</i>	[23]
	<b>17</b>	Nonacosanoic acid, 2-(4-hydroxyphenyl)ethyl ester	Stem barks	<i>B. africana</i>	[18]
	<b>18</b>	Zeyherol	Stem wood	<i>B. digitalis</i>	[21]
<b>III-Lignans</b>					
	<b>19</b>	(+)-Lyoniresinol 3-α-O-β-D-glucopyranoside	Aerial part	<i>B. unguis-cati</i>	[14]
<b>IV-Flavonoids</b>					
	<b>20</b>	Brachydin A	Root	<i>B. brachypoda</i>	[6, 24]
	<b>21</b>	Brachydin B			
	<b>22</b>	Brachydin C			
	<b>23</b>	Brachydin D	Root	<i>B. brachypoda</i>	[6]
	<b>24</b>	Brachydin E			
	<b>25</b>	Brachydin F			
	<b>26</b>	Brachydin G			
	<b>27</b>	Brachydin H			
	<b>28</b>	Brachydin I			
	<b>29</b>	Brachydin J			
	<b>30</b>	Cirsimarin	Aerial part	<i>B. unguis-cati</i>	[14]
	<b>31</b>	4'-Hydroxywogonin	Leaf	<i>B. callistigoides</i>	[15]

**Table 2:** A list of previously reported compounds from genus "Bignonia" (with different synonyms as shown in Table 1) (cont.).

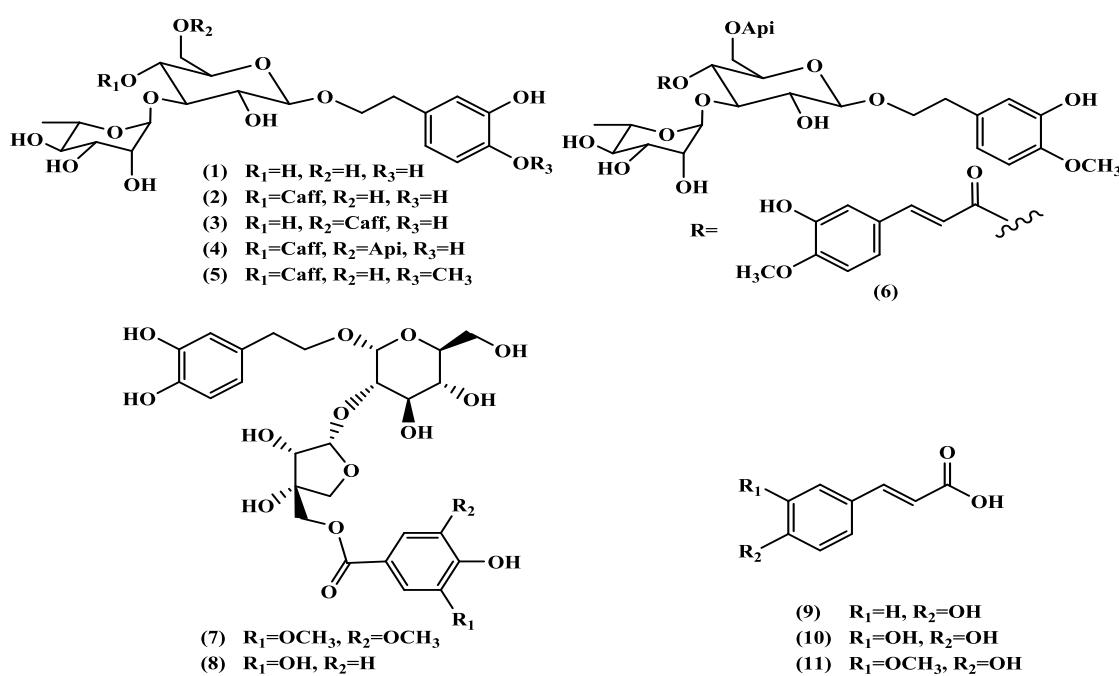
Classification	No.	Item	Part used	Source	Ref.
	32	Acacetin	Leaf	<i>B. callistigooides</i>	[15]
			Leaf	<i>B. chica</i>	[25]
	33	Galangustin	Leaf	<i>B. callistigooides</i>	[15]
	34	7,8-Dimethoxyflavone	Leaf	<i>B. aesculifolia</i>	[26]
	35	5,6,7,8-Tetramethoxyflavone			
	36	Alnetin	Leaf	<i>B. aesculifolia</i>	[27]
	37	Gardenin B			
	38	Chrysin 7-methyl ether			
	39	3',4'-Dihydroxy-5,6,7-trimethoxyflavone	Leaf	<i>B. brachypoda</i>	[28]
	40	Cirsiliol			
	41	Cirsimaritin	Aerial part	<i>B. unguis-cati</i>	[14]
			Leaf	<i>B. brachypoda</i>	[28]
	42	Hispidulin	Leaf	<i>B. brachypoda</i>	[28]
	43	Chrysin	Leaf	<i>B. samydoides</i>	[29]
	44	Carajuflavone	leaf	<i>B. chica</i>	[30]
	45	Luteolin	Leaf	<i>B. stans</i>	[19]
			leaf	<i>B. chica</i>	[30]
	46	Sorbarin	Leaf	<i>B. elegans</i>	[31, 32]
	47	Triumboidin	Leaf	<i>B. elegans</i>	[32]
	48	Apigenin	Leaf	<i>B. stans</i>	[19]
	49	Chrysoeriol			
	50	Corymboside	Leaf	<i>B. unguis-cati</i>	[23]
	51	Vicenin-2	Leaf	<i>B. unguis-cati</i>	[12, 23]
	52	6-Methoxy acacetin 7-O-glucoside	Aerial part	<i>B. unguis-cati</i>	[12]
	53	8-Methoxy acacetin 7-O- glucoside			
	54	Acacetin 7-O-glucosyl-8-C-rhamnosyl-3-O- $\alpha$ -arabinofuranoside			
	55	4'-O-Methyl scutellarin 6-O-apiosyl galactoside			
	56	4'-Methyl-6-methoxy kaempferol 7-O-8-C diglucoside			
	57	6-Methoxy Apigenin 7-O- glucoside			
	58	Quercitrin	Aerial part	<i>B. unguis-cati</i>	[12]
			Leaf	<i>B. unguis-cati</i>	[23]
	59	Luteolin 7,4'-dimethyl ether	Leaf	<i>B. aesculifolia</i>	[27]
	60	Alpinetine	Leaf	<i>B. triplinervia</i>	[33]
	61	Carajurin	Leaf	<i>B. chica</i>	[25]
	62	6,7,4'-Trihydroxy-5-methoxy-flavylium			
	63	6,7,3',4'-Tetrahydroxy-5-methoxy-flavylium			
	64	6,7,3'-Trihydroxy-5,4'-dimethoxy-flavylium			
<b>V-Coumarins</b>					
	65	6-Methoxymellein	Root and bark	<i>B. africana</i>	[34]
	66	Kigelin			
	67	6-Demethylkigelin			
<b>VI-Sterols</b>					
	68	$\beta$ -Sitosterol	Heartwood	<i>B. adenophylla</i>	[35]
			Stem bark	<i>B. africana</i>	[18]
			Leaf	<i>B. samydoides</i>	[29]
			Root and bark	<i>B. africana</i>	[34]
			Leaf	<i>B. unguis-cati</i>	[23]
	69	$\beta$ -Sitosterol 3-O- $\beta$ -D-glucopyranoside	Aerial part	<i>B. unguis-cati</i>	[36]
	70	Stigmasterol	Leaf	<i>B. unguis-cati</i>	[23]
			Root and bark	<i>B. africana</i>	[34]
			Leaf	<i>B. samydoides</i>	[29]
<b>VII-Triterpenes</b>					
	71	3 $\beta$ -estearioxy-olean-12-ene	Root	<i>B. brachypoda</i>	[37]
	72	Oleanolic acid	Stem bark	<i>B. africana</i>	[18]
			Leaf	<i>B. triplinervia</i>	[33]
			Leaf	<i>B. chica</i>	[25]
	73	Pomolic acid	Stem bark	<i>B. africana</i>	[18]
			Leaf	<i>B. triplinervia</i>	[33]
	74	2 $\beta$ ,3 $\beta$ ,19 $\alpha$ -Trihydroxy-urs-12-en-28-oic acid	Stem bark	<i>B. africana</i>	[18, 22]
	75	$\beta$ -Friedelinol			
	76	$\beta$ -Amyrin	Heartwood	<i>B. adenophylla</i>	[35]
			Stem bark	<i>B. unguis-cati</i>	[36]
	77	3 $\beta$ ,16 $\alpha$ -Dihydroxy-urs-12-ene	Leaf	<i>B. samydoides</i>	[29]
	78	Erythrodiol			
	79	Ursolic acid	Aerial part	<i>B. unguis-cati</i>	[14]
			Leaf	<i>B. triplinervia</i>	[33]
			Leaf	<i>B. samydoides</i>	[29]

**Table 2:** A list of previously reported compounds from genus "Bignonia" (with different synonyms as shown in Table 1) (cont.).

<b>Classification</b>	<b>No.</b>	<b>Item</b>	<b>Part used</b>	<b>Source</b>	<b>Ref.</b>
	<b>80</b>	Uvaol	Leaf	<i>B. samydoides</i>	[29]
	<b>81</b>	Lupeol	Leaf	<i>B. samydoides</i>	[29]
	<b>82</b>	Quinovic acid	Leaf	<i>B. unguis-cati</i>	[23]
	<b>82</b>	Quinovic acid	Aerial part	<i>B. unguis-cati</i>	[36]
<b>VIII-Diterpenes</b>					
	<b>83</b>	Phytol	Leaf	<i>B. africana</i>	[38]
	<b>84</b>	3-Hydro-4,8-phytene			
<b>IX-Unsaturated fatty acids</b>					
	<b>85</b>	(9Z,12Z)-Methyl octadeca-9,12-dienoate	Leaf	<i>B. africana</i>	[38]
	<b>86</b>	Linoleic acid	Leaf	<i>B. callistigoides</i>	[39]
	<b>87</b>	Linolenic acid			
<b>X-Saturated fatty acids</b>					
	<b>88</b>	Stearic acid	Leaf	<i>B. callistigoides</i>	[39]
	<b>89</b>	Palmitic acid			
<b>XI-Xanthones</b>					
	<b>90</b>	Mangiferin	Leaf	<i>B. patellifera</i>	[7]
	<b>91</b>	3'-O-p-Hydroxybenzoylmangiferin			
	<b>92</b>	3'-O-Trans-coumaroylmangiferin			
	<b>93</b>	6'-O-Trans-coumaroylmangiferin			
	<b>94</b>	3'-O-Trans-cinnamoylmangiferin			
	<b>95</b>	3'-O-Trans-caffeoymangiferin			
	<b>96</b>	3'-O-Benzoylmangiferin			
<b>XII-Quinones</b>					
	<b>97</b>	Lapachol	Stem wood Stem bark Roots Whole plant Stem and fruit Aerial part Stem wood roots and bark Stem heart wood	<i>B. adenophylla</i> <i>B. africana</i> <i>B. aesculifolia</i> <i>B. unguis-cati</i> <i>B. africana</i> <i>B. unguis-cati</i> <i>B. digitalis</i> <i>B. africana</i> <i>B. adenophylla</i>	[17] [18] [40] [23] [20] [36] [21] [34] [17]
	<b>98</b>	Peshawaraquinone			
	<b>99</b>	Methyl-1,2-dihydroxy-2-(3-methylbut-2-en-1-yl)-3-oxo-2,3-dihydro-1H-indene-1-carboxylate			
	<b>100</b>	$\alpha$ -Lapachone			
	<b>101</b>	Dehydro $\alpha$ -lapachone	Stem bark Stem and fruit	<i>B. africana</i> <i>B. africana</i>	[18] [20]
	<b>102</b>	3,4-Dihydroxy-2-(3-methylbut-2-en-1-yl)-3,4-dihydroronaphthalen-1(2H)-one	Root	<i>B. aesculifolia</i>	[40]
	<b>103</b>	$\beta$ -Lapachone			
	<b>104</b>	Dilapachone	Heartwood	<i>B. adenophylla</i>	[35]
	<b>105</b>	Adenophyllone			
	<b>106</b>	Tecomaquinone I			
	<b>107</b>	Dehydro iso $\alpha$ -lapachone			
	<b>108</b>	Tectol			
	<b>109</b>	2-Acetylfur-1,4-naphthoquinone	Stem bark	<i>B. africana</i>	[18]
	<b>110</b>	Kigelinol	Stem bark Root and stem bark	<i>B. africana</i> <i>B. africana</i>	[18] [41]
	<b>111</b>	Isokigelinol	Root and stem bark	<i>B. africana</i>	[41]
	<b>112</b>	Isopinnatal	Root and stem bark	<i>B. africana</i>	[41]
	<b>113</b>	Norviburtina	Root and fruit	<i>B. africana</i>	[20]
	<b>114</b>	2-(1-Hydroxyethyl)-naphtho[2,3-b]furan-4,9-quinone	Root bark	<i>B. africana</i>	[42]
	<b>115</b>	Kigelinone	Root and stem bark	<i>B. africana</i>	[41]
	<b>116</b>	Kojic acid	Root and fruit Stem bark	<i>B. africana</i> <i>B. africana</i>	[20] [18]
<b>XIII- Limonoids</b>					
	<b>117</b>	1-O-Deacetyl-2 $\alpha$ -methoxykhayanolide	Whole plant	<i>B. africana</i>	[43]
	<b>118</b>	Deacetylkhayanolide E			
	<b>119</b>	1-O-Deacetyl-2 $\alpha$ -hydroxykhayanolide E			
	<b>120</b>	Kigelianolide			
	<b>121</b>	Khayanolide B			
<b>XIV- Iridoids</b>					
	<b>122</b>	Theviridoside	Stem	<i>B. crucigera</i>	[5]
	<b>123</b>	6'-O-Cyclopropanoyl theviridoside			
	<b>124</b>	10-O-p-Hydroxybenzoyl theviridoside			

**Table 2:** A list of previously reported compounds from genus "Bignonia" (with different synonyms as shown in Table 1) (cont.).

Classification	No.	Item	Part used	Source	Ref.
	125	10-O-Vanillyltheviridosid			
	126	7-Hydroxyviteoid II	Fruit	<i>B. africana</i>	[44]
	127	7-Hydroxy eucommic acid			
	128	7-Hydroxy-10-deoxyeucommic acid			
	129	10-Deoxyeucommiol			
	130	Jiofuran			
	131	Jioglutolide			
	132	1-Dehydroxy-3,4-dihydroaucubigenin			
	133	Des- <i>p</i> -hydroxy benzoyl kisasagenol B			
	134	Ajugol			
	135	6-Trans-caffeooyl ajugol			
	136	Vermoside	Bark and fruit	<i>B. africana</i>	[16, 44]
	137	Specicoside	Stem bark	<i>B. africana</i>	[16, 22]
	138	Minecoside	Bark	<i>B. africana</i>	[45]
	139	Lamiide	Leaf	<i>B. elegans</i>	[31, 46]
	140	Pseudocalymmoside	Whole plant	<i>B. elegans</i>	[46]
	141	Durantosid II			
	142	Eleganoside A	Leaf	<i>B. elegans</i>	[32]
	143	Eleganoside B			
	144	Eleganoside C			
	145	Stansioside	Leaf	<i>B. stans</i>	
	146	Plantarenaloside			
	147	5-Deoxystansioside			
<b>XV- Alkaloids and nitrogenous compounds</b>					
	148	4-Hydroxytecomanine	Fruit	<i>B. stans</i>	[48]
	149	Tecomanine (Tecomine)	Leaf	<i>B. stans</i>	[49, 50]
	150	5β-Hydroxyskitanthine	Fruit	<i>B. stans</i>	[48]
			Leaf	<i>B. stans</i>	[50]
	151	4-Noractnidin	Leaf	<i>B. stans</i>	[49, 50]
	152	Boschniakine			
	153	Tecostanine	Leaf	<i>B. stans</i>	[50]
	154	Allantoin	Leaf	<i>B. unguis-cati</i>	[23]
	155	Pheophorbide a	Leaf	<i>B. elegans</i>	[31]
	156	Indole	Leaf	<i>B. chica</i>	[51]
	157	Tryptophan		<i>B. stans</i>	[52]
	158	Tryptamine			
	159	Skatole			
<b>XVI- Dipeptides</b>					
	160	N(N'-Benzoyl-S*-phenylalaninyl)-S*-phenylalaninol benzoate	Root	<i>B. digitalis</i>	[53]

**Figure 1:** Compounds isolated from genus "Bignonia".

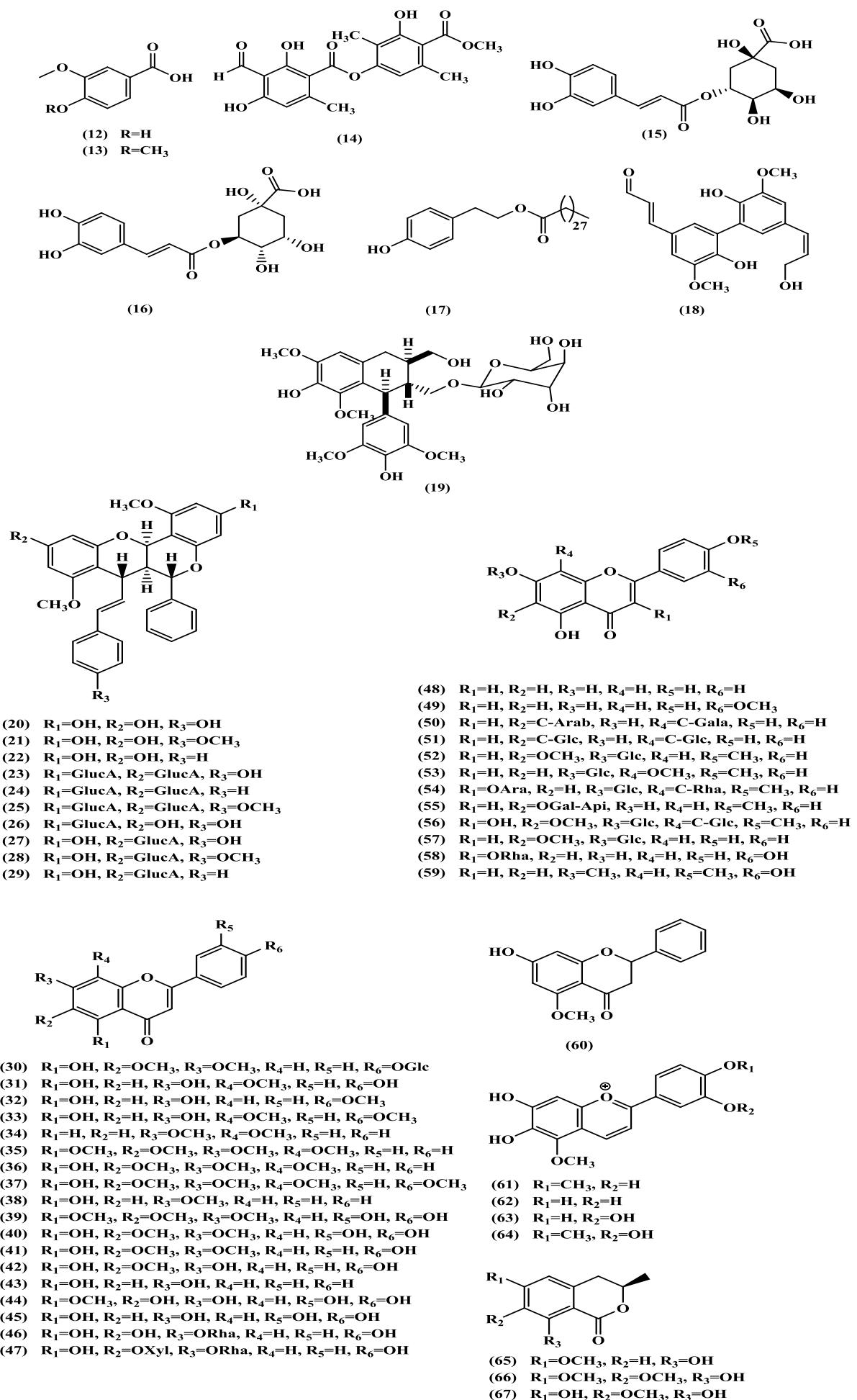
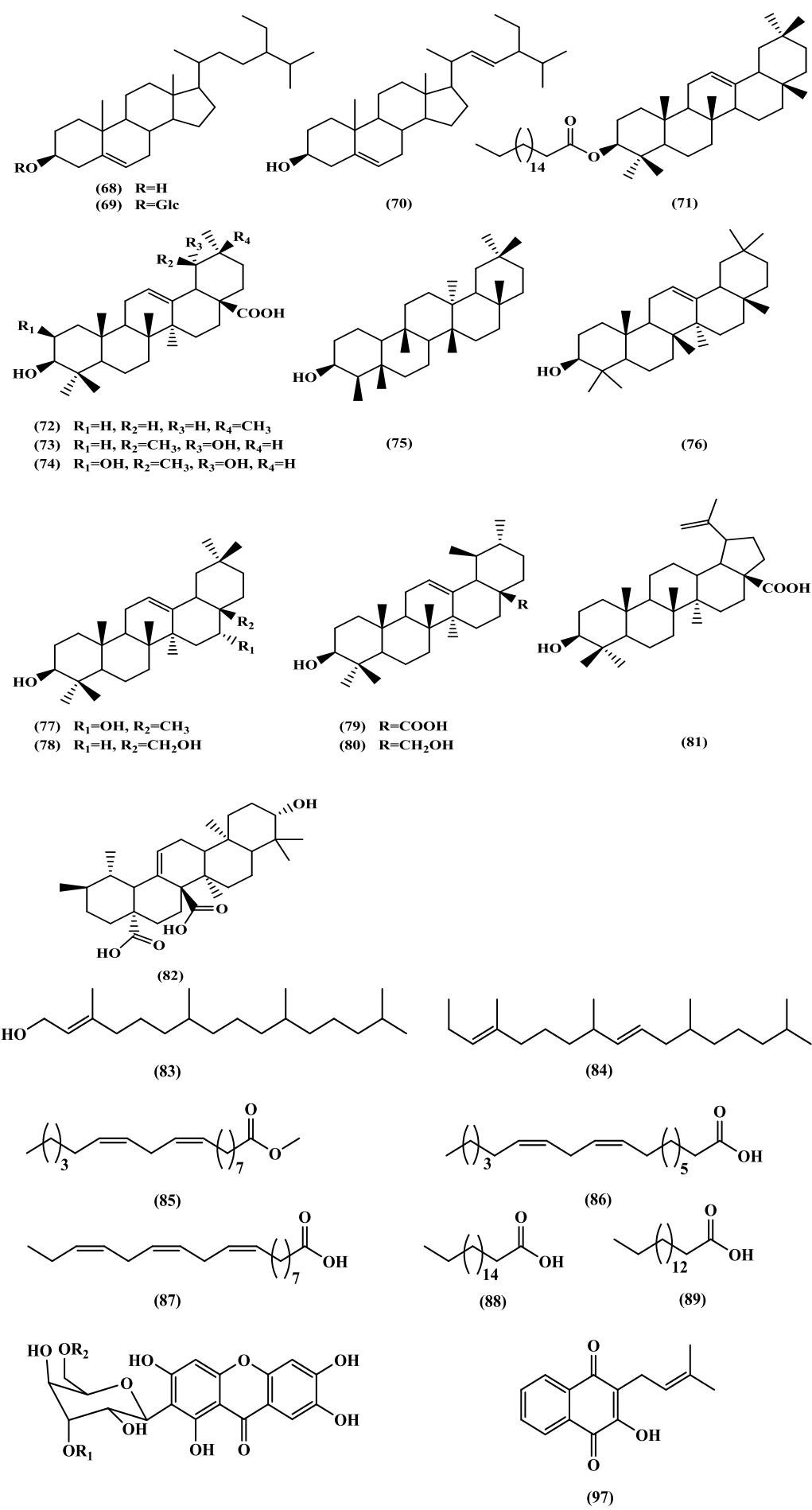


Figure 1: Compounds isolated from genus "Bignonia" (cont.).

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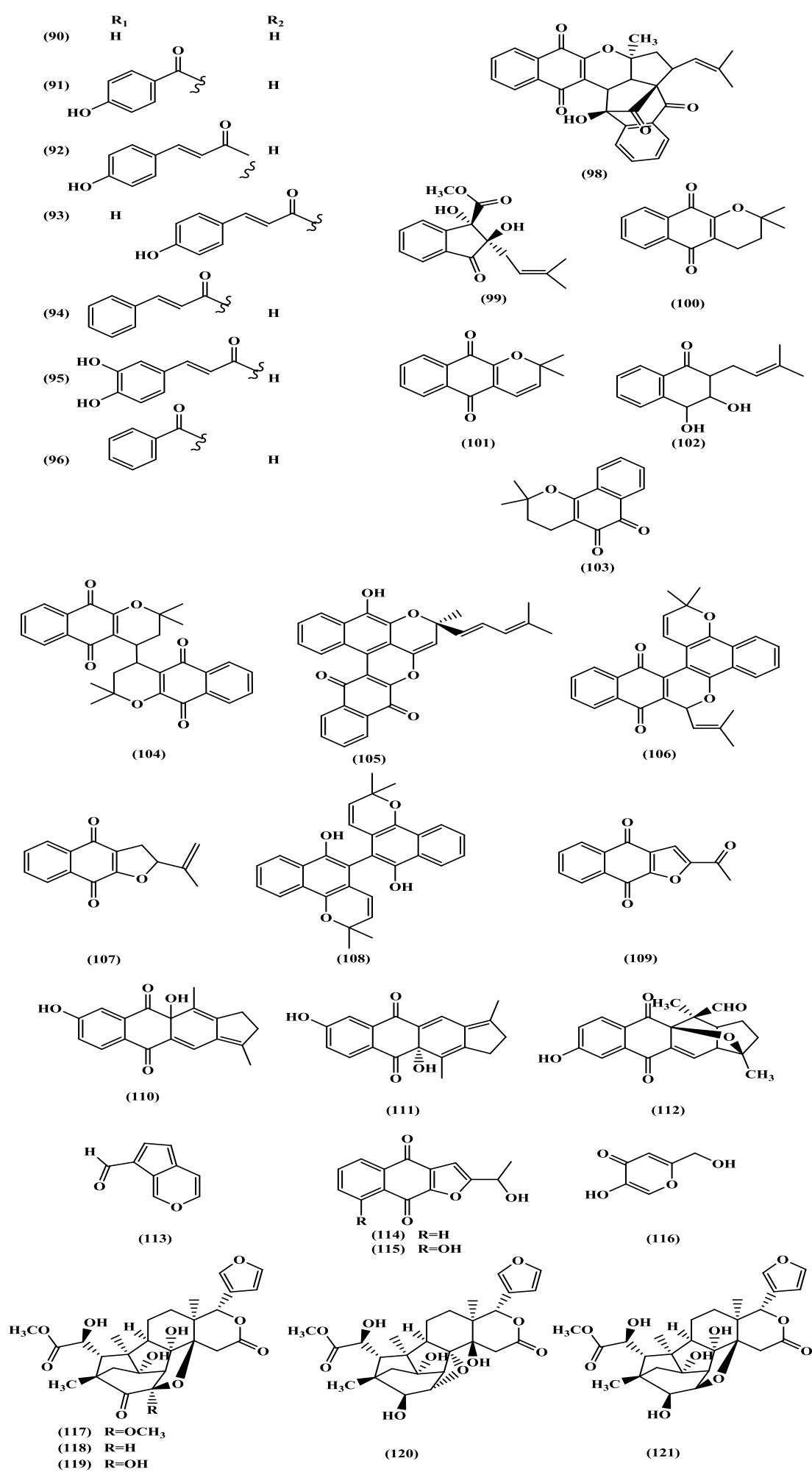


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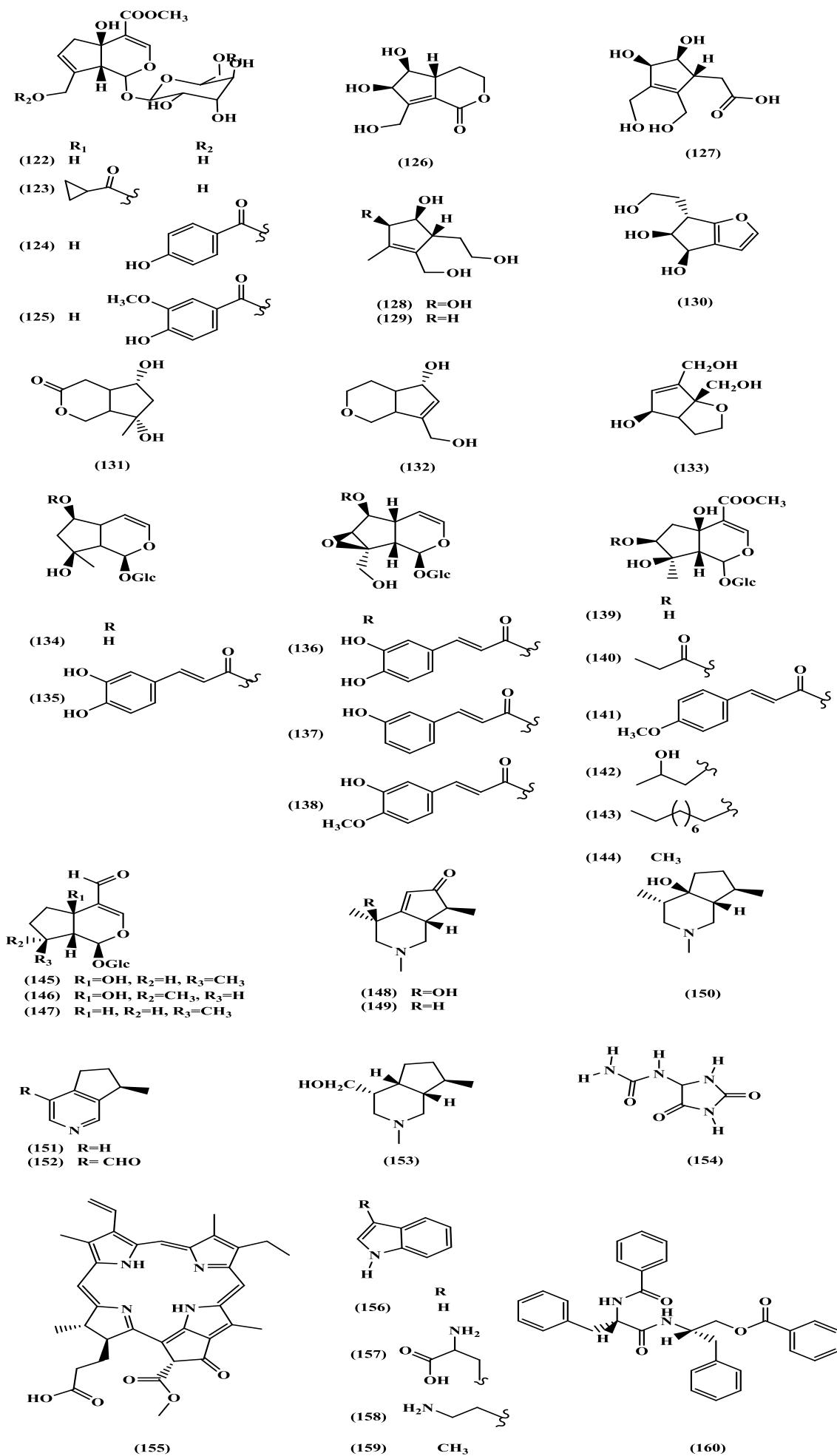


Figure 1: Compounds isolated from genus "Bignonia" (cont.).

### 3.3. Quinones

Quinones are widely distributed in *Bignonia* species, as lapachol,  $\alpha$ -lapachone and dehydro  $\alpha$ -lapachone in *B. africana* and *B. adenophylla* [17, 18].

### 4. Biological activities of genus "Bignonia"

*Bignonia* species exhibited various biological activities including; cytotoxic [54] antiprotozoal [51], analgesic [55], insect repellent [15], anti-inflammatory [37] and cholinergic activities [43] as demonstrated in (Table 3).

**Table 3:** A list of different biological activities of genus "Bignonia" (with different synonyms as shown in Table 1).

Biological activity	Plant name /part used	Extract, fraction or compound	Method/ Result	Ref.
<b>I- Anti-insect activity</b>				
<i>B. callistigoides</i> Leaf	Hydrolysed methanol extract 4'-Hydroxywogonin Acacetin Galangustin		They showed different settling inhibition activity against <i>Myzus persicae</i> and <i>Rhopalosiphum padi</i> , agricultural aphids, in which the hydrolyzed methanol extract was active only against <i>M. persicae</i> , while acacetin had activity only against <i>R. padi</i> (PI=0.3±0.1), Galangustin had activity against both aphids (PI=0.4±0.1) for <i>M. persicae</i> and 0.8±0.1 for <i>R. padi</i> and 4'-Hydroxywogonin had no any activity. Stearic acid, linoleic acid and linolenic acid inhibit <i>R. padi</i> settling, while palmitic acid was inactive.	[15]
<i>B. africana</i> Bark	Palmitic acid Stearic acid Linoleic acid Linolenic acid Aqueous extract		Its mollucidal effect was tested on Nile tilapia, <i>Oreochromis niloticus</i> (L.), using continuous aeration over a period of 96 h result in eventual fatigue and eventual death.	[56]
<b>II- Steroidogenic enzymes effect</b>				
<i>B. capensis</i>	Ethanol extract		It showed rhythm in estrus cyclicity and ovarian weights similar to controls in letrozole-induced polycystic ovarian model.	[57]
<b>III- Antimicrobial activity</b>				
<i>B. adenophylla</i> Leaf and seed	Aqueous and methanol extracts		They were tested using the disc diffusion method, in which the aqueous extract of the seeds showed maximum potency against <i>Staphylococcus aureus</i> and <i>Staphylococcus epidermidis</i> , while the methanol extract showed maximum activity against <i>Escherichia coli</i> , on the other hand, the methanol extract of the leaves showed the maximum activity against <i>S. aureus</i> .	[58]
<i>B. africana</i> Stem bark	p-Coumaric acid Caffeic acid Kojic acid		They showed varied anti-candidal activity against four <i>Candida albicans</i> strains (ATCC26, ATCC12C, ATCCP37039 and ATCCP37037) through agar diffusion method and microbroth dilution technique, in which kojic acid was the most potent.	[18]
<i>B. cuprea</i> Leaf	Hydroethanol extract		It showed activity against <i>Helicobacter pylori</i> and moderate activity against <i>Enterococcus faecalis</i> in broth microdilution method.	[59]
<i>B. aesculifolia</i> Root	Ethanol extract		It showed no antifungal activity against subcutaneous fungi, <i>Sporothrix schenckii</i> and <i>Fonsecaea pedrosoi</i> using 100 µg/ml.	[60]
<i>B. callistigoides</i> Bark	Ethanol extract		It showed no antifungal activity against sc. fungi, <i>S. schenckii</i> and <i>F. pedrosoi</i> using 100 µg/ml.	[60]
<i>B. brachypoda</i> Leaf	Cirsimarin, Cirsiliol 3',4'-Dihydroxy-5,6,7-trimethoxyflavone Hispidulin		They showed antifungal activity against <i>Cladosporium sphaerospermum</i> spores by direct bioautography method on TLC plates, but hispidulin is the less active one.	[28]
<i>B. africana</i> Root and fruit	Methanol extract Kigelinone Isopinnatal Dehydro- $\alpha$ -lapachone Caffeic acid <i>p</i> -Coumaric acid Ferulic acid		It showed antibacterial activity only against the Gram-positive organisms. They showed various antibacterial and antifungal activities against different microorganisms, in which kigelinone was the most active one, followed with the caffeic acid.	[20]
<b>IV- Cholinergic activity</b>				
<i>B. africana</i> Whole plant	1-O-Deacetyl-2 $\alpha$ -methoxy khyanolide Deacetylkhyanolide E 1-O-Deacetyl-2 $\alpha$ -hydroxy Khyanolide E Kigelianolide Khyanolide B		They had weak inhibition activities against acetylcholinesterase and butyrycholinesterase.	[43]

**Table 3:** A list of different biological activities of genus "Bignonia" (with different synonyms as shown in Table 1) (cont.).

Biological activity	Plant name /part used	Extract, fraction or compound	Method/ Result	Ref.
<b>V- Antiprotozoal activity</b>				
<i>B. cuprea</i> Leaf	Petroleum ether fraction Hexane fraction Chloroform fraction EtOAc fraction Pheophorbide a		They showed various activities against <i>Trypanosoma cruzi</i> on different stages, but the chloroform fraction was the best one. While, pheophorbide a activity was tested in the presence and absence of light, in which its effect increased against the main infective forms of <i>Trypanosoma cruzi</i> (tryomastigotes and amastigotes) in the presence of light.	[51]
<i>B. brachypoda</i> Root	Nonpolar fraction Brachydin A Brachydin B Brachydin C		The Nonpolar fraction was active against <i>T. cruzi</i> , while brachydin B, brachydin C showed selective inhibition activity, in which they inhibited the parasite invasion process and its intracellular development in host cells with similar potencies to benznidazole. Additionally, brachydin B decreased the blood parasitemia of <i>T. cruzi</i> -infected mice.	[24]
<i>B. africana</i> Stem bark	Specicoside $2\beta,3\beta,19\alpha$ -Trihydroxy urs-12-en-28-oic acid Atranorin <i>p</i> -Hydroxy cinnamic acid		The antiplasmodial activity was evaluated against the multidrug-resistant W2mef strain of <i>Plasmodium falciparum</i> by the parasite lactate dehydrogenase assay, in which specicoside, $2\beta,3\beta,19\alpha$ -trihydroxyurs-12-en-28-oic acid and atranorin were significantly active.	[22]
<i>B. patellifera</i> Leaf	Mangiferin $3'-O-p$ -Hydroxybenzoylmangiferin $3'-O$ -trans coumaroylmangiferin		They were active <i>in vitro</i> against the chloroquine-sensitive <i>P. falciparum</i> 3D7 clone.	[7]
<i>B. patellifera</i> Leaf	Ethanol extract Ursolic acid Alpinetine Oleanolic acid		They showed trypanocidal activity against <i>T. cruzi</i> , in which ursolic acid was four times more active than oleanolic acid, while alpinetine was inactive.	[33]
<i>B. africana</i> Bark	Verminoside Specioside Minecoside		Their antiamoebic were evaluated <i>in vitro</i> against <i>Entamoeba histolytica</i> HK-9 strain, in which verminoside was the most active one followed by specioside.	[16]
<i>B. africana</i> Root and stem bark	2-(1-Hydroxyethyl)naphtho[2,3- <i>b</i> ]furan-4,9-quinone Isopinnatal Kigelinol Isokigelinol		They showed different antitrypanosomal activity against <i>T. brucei brucei</i> and <i>T. brucei rhodesiense</i> , in which 2-(1-hydroxyethyl)naphtho[2,3- <i>b</i> ]furan-4,9-quinone was the most potent.	[41]
<i>B. unguis-cati</i> Whole plant	Ethanol extract Pet. ether extract EtOAc extract		They did not show antiprotozoal activity against <i>Leshmania</i> sp. or <i>T. cruzi</i> .	[61]
<b>VI- Analgesic activity</b>				
<i>B. brachypoda</i> Root	Ethanol extract		It showed clear potency in formalin and acetic acid induced writhing tests activity.	[62]
<i>B. hymenaea</i> Leaf	Aqueous extract		It showed significant activity in acetic acid-induced writhing, formalin (paw licking test) and tail flick testes with a possible involvement of central mechanism and adenosine system test in mice.	[55]
<i>B. africana</i> Stem bark	Ethanol extract		It tested by using acetic acid induced mouse writhing, in which it decreased the number of writhes in a dose dependent manner while, in hot plate reaction time, it showed insignificant elongation of the reaction time.	[63]
<b>VII- Anti-inflammatory activity</b>				
<i>B. cuprea</i> Leaf	Ethanol and aqueous extracts		They reduced the inflammation in murine sponge model by decreasing neutrophil accumulation and hemoglobin content in the sponge implants without altering the level of cytokines.	[64]
<i>B. brachypoda</i> Root	$3\beta$ -Estearioxy-olean-12-ene		It showed potent anti-inflammatory activity by using carrageenan-induced paw edema, formalin and hot plate tests.	[37]
<i>B. brachypoda</i> Root	Ethanol extract		It showed marked anti-inflammatory activity against carrageenan-induced paw edema, peritonitis and fibrovascular tissue growth, which induced by subcutaneous cotton pellet implantation.	[62]
<i>B. unguis-cati</i> Aerial part	Ethanol extract		The ethanol extract was more potent extract than chloroform in reducing the carrageenan-induced paw edema.	[12]
<i>B. africana</i> Stem bark	Chloroform extract		It showed a significant dose dependent carrageenan induced paw edema inhibition between the 2 <sup>nd</sup> and 5 <sup>th</sup> h.	[63]
<i>B. brachypoda</i> Leaf	$3',4'$ -Dihydroxy-5,6,7-trimethoxy flavone		It inhibited arachidonate 5-lipoxygenase enzyme.	[28]
<i>B. cuprea</i> Leaf	Carajurin		It was studied by investigation the NF- $\kappa$ B and NF-AT assays, in which it inhibited NF- $\kappa$ B, but not NF-AT at 500 mM.	[25]
<b>VIII- Antidiarrheal activity</b>				
<i>B. africana</i> Leaf	Aqueous extract		It was examined by using castor oil-induced animal model. It decreased the fecal output by reducing nicotine induced contractions.	[65]

**Table 3:** A list of different biological activities of genus "Bignonia" (with different synonyms as shown in Table 1) (cont.).

Biological activity	Plant name /part used	Extract, fraction or compound	Method/ Result	Ref.
<b>IX- Cytotoxic activity</b>				
<i>B. africana</i>	Seed oil		It inhibited significantly dose-dependently Caco-2 cell growth compared to HEK-293 cell growth.	[54]
Seed				[12]
<i>B. unguis-cati</i>	Ethanol extract		It was the most effective sample on lung cell line.	
Aerial part				
<i>B. africana</i>	DCM extracts		The extracts were tested <i>in vitro</i> , using sulphorhodamine B assay against cultured melanoma and other cancer cell lines, in which norviburtinal was found to be the most active compound with little selectivity, while isopinnatal also showed some cytotoxic activity.	[66]
Stem bark and fruit	Norviburtinal			
	Isopinnatal			
	$\beta$ -Sitosterol			
<i>B. africana</i>	Aqueous extract		They were investigated for their growth inhibitory effects against four melanoma cell lines and Caki-2 using two different assays.	[67]
Stem bark and fruit	Ethanol extract			
	DCM extract		The DCM extract of the stem bark and lapachol had significant dose-dependent inhibitory activity, while the extract was less active after an hour exposure.	
	Lapachol		The chemosensitivity of the melanoma cell lines to the stem bark was greater than that seen for the renal adenocarcinoma line. In which sensitivity to lapachol was similar amongst the five cell lines.	
<b>X- Sleep induction</b>				
<i>B. africana</i>	Ethanol extract		It reduced the duration of sleeping time in dose-dependent manner in barbiturate-induced sleeping.	[68]
Stem bark				
<b>XI- Male fertility</b>				
<i>B. africana</i>	Aqueous extract		It showed ability to reverse <i>Carica papaya</i> induced testicular damage, if administered within a certain window period such as:	[69]
Bark			*If administrated within 4 weeks of treatment with <i>C. papaya</i> reversed the deleterious effects on semen parameters.	
			*If given after 10 weeks, the damage remains unreversed.	
<b>XII- Anti-obesity</b>				
<i>B. hymenaea</i>	Aqueous extract		It reduced significantly the plasma LDL-cholesterol and triglycerides comparing to the atorvastatin in diet-induced hypercholesterolemia.	[70]
Leaf				
<b>XIII- Heart protection</b>				
<i>B. stans</i>	70% Ethanol extract		It prevented the reduction of the antioxidants and retarded elevation of cardiac damage markers in isoproterenol-induced myocardial infarction rats.	[71]
Flower				
<b>XIV- Hepatic and renal protection</b>				
<i>B. hymenaea</i>	Aqueous extract		The aqueous extract caused an increase in Glucose, creatinine and albumin parameters. While, urea, aspartate aminotransferase and alanine aminotransferase values were decreased with (50 and 100 mg/kg) and increased with dose of 200 mg/kg. But, ethanol extract caused an increase in this parameter to the doses used. In addition to, their ability to cause leukopenia.	[72]
Leaf	Ethanol extract			
<i>B. stans</i>	EtOAc extract		It significantly protected rat kidneys in gentamicin-induced nephrotoxicity albino rats.	[73]
Flower				
<b>XV- Gastric protection</b>				
<i>B. Brachypoda</i>	Hydroethanol extract		It protected significantly the stomach against absolute ethanol, depletion of (glutathione and nitric oxide), non-steroidal anti-inflammatory drugs, pylorus ligation and acetic acid-induction gastric ulcer in rats.	[6]
Root				
<b>XVI- Protection against light</b>				
<i>B. cuprea</i>	A crude extract		The crude extract and chloroform fraction showed photochemoprotective activity in protecting L929 fibroblasts against both UVA and UVB-induced cell damage through scavenging mechanisms.	[74]
Leaf	Hexane fraction			
	EtOAc fraction			
	Chloroform fraction			
<i>B. unguis-cati</i>	Petroleum ether fraction			
Aerial part	Chloroform fraction			
	EtOAc fraction			
	<i>n</i> -Butanol fraction			
<i>B. africana</i>	Aqueous fraction			
Leaf	Hexane fraction			
	EtOAc fraction			
	Methanol fraction			
<i>B. patellifera</i>	Methanol extract			
Leaf	Mangiferin			
	3'- <i>O</i> - <i>p</i> -Hydroxy benzoylmangiferin			

**Table 3:** A list of different biological activities of genus "Bignonia" (with different synonyms as shown in Table 1) (cont.).

Biological activity	Plant name /part used	Extract, fraction or compound	Method/ Result	Ref.
<i>B. cuprea</i>	Leaf	3'-O-Trans-coumaroylmangiferin 6'-O-Trans-coumaroylmangiferin 3'-O-Trans-cinnamoylmangiferin 3'-O-Trans-caffeoymangiferin 3'-O-Benzoyl mangiferin		
<i>B. cuprea</i>	Leaf	Methanol/0.3% citric acid extract	It exhibited moderate scavenging activity on DPPH assay.	[75]
<i>B. crucigera</i>	Stem	Methanol extract Verbascoside Isoverbascoside Forsythoside B Jionoside D Leucosceptoside B Carajurin	The extract exhibited activity against DPPH by using rapid TLC tests. The isolated compounds showed potent radical-scavenging activity against DPPH.	[5]
<i>B. cuprea</i>	Leaf	6,7,4'-Trihydroxy-5-methoxy-flavylium 6,7,3',4'-Tetrahydroxy- 5-methoxy-flavylium 6,7,3'-Trihydroxy-5, 4'-dimethoxy flavylium	They exhibited no activity on DPPH TLC analysis.	[25]
<b>XVIII- Wound healing activity</b>				
<i>B. cuprea</i>	Whole plant	Lyophilized extract	It improved collagen organization and increased the quantity of dermatan sulfate on the 14 <sup>th</sup> day of the tendon healing.	[76]
<i>B. cuprea</i>	Leaf	Methanol/0.3% citric acid extract	It exhibited efficient wound healing activity on both <i>in vivo</i> and <i>in vitro</i> using fibroblast growth and collagen production stimulation assays.	[75]
<b>XIX- Anti-diabetic activity</b>				
<i>B. stans</i>	Leaf	Hydroalcohol extract Flavone fractions Chrysoeriol Apigenin Luteolin Verbascoside	The mixtures of (chrysoeriol and apigenin) and (chrysoeriol and luteolin) showed significant lipase inhibitory activity <i>in vitro</i> by indirect spectrophotometric method type 2 diabetes mellitus.	[19]
<i>B. stans</i>	Leaf	Aqueous extract	It evaluated <i>in vitro</i> on the adipogenesis and 2-NBDglucose uptake in insulin-sensitive and insulin-resistant murine 3T3-F442A and human subcutaneous adipocytes. It stimulated 2-NBDG uptake in a concentration-dependent manner.	[77]
<i>B. stans</i>	Leaf	Aqueous extract	It displayed <i>in vitro</i> a dose-dependent inhibition of glucose release from starch.	[78]
<i>B. stans</i>	Leaf	Tecomine Boschniakine 5β-Hydroxyskitanthine Tecostanine	Tecomine only affected markedly on glucose uptake <i>in vitro</i> glucose uptake in white adipocytes, while none of them was active <i>in vivo</i> on albino mice, a genetic model of type II diabetes.	[50]

## 5. Conclusion

The literature survey of different *Bignonia* species revealed the presence 160 compounds of different classes of phytoconstituents as flavonoids (45 compound), iridoids (26) and quinones (20), which are the most abundant among the various species of *Bignonia*, followed by triterpenes (12), phenolic acid derivatives (12) and alkaloids (12). Besides, 5 limonoids, 7 xanthones and 6 phenylethanoids. Furthermore, 9 compounds belong to different classes of secondary metabolites e.g., coumarins (3), sterols (3) and unsaturated fatty acids (3). Finally, saturated fatty acids (2) and diterpenes (2). This review showed the diversity of biological activities exhibited by *Bignonia* plants as cytotoxic as *B. unguis-cati* and *B. africana*, antioxidant as *B. cuprea*, *B. patellifera*, *B. unguis-cati* and *B. africana*, antileishmanial as *B. unguis-cati*, antitrypanosomal as

*B. cuprea*, *B. patellifera* and *B. brachyboda* and antimarial as *B. africana* and *B. patellifera*. Besides, antimicrobial of the different species, analgesic as *B. hymenaea*, *B. brachyboda* and *B. africana*. In addition to the anti-inflammatory of *B. cuprea*, *B. brachyboda* *B. unguis-cati* and *B. africana*. Also, some plants of *Bignonia* exhibited wound healing potential, antidiabetic activity, sleep induction, gastroprotective and anti-obesity activities.

Based on, the present review *Bignonia* plants need further investigation to study their safety and confirm their application in folk medicine. Although, *Bignonia* genus comprises of 31 species, only 15 species reported in the literature, while the others uninvestigated neither phytochemically nor biologically. Moreover, *Bignonia* plants need further studies as they are considered a good source of bioactive natural products.

## References

- [1] Rahmatullah M, Samarrai W, Jahan R, Rahman S, Sharmin N, Emdad Ullah Miajee ZUM, et al. An ethnomedicinal, pharmacological and phytochemical review of some Bignoniaceae family plants and a description of Bignoniaceae plants in folk medicinal uses in Bangladesh. *Advances in Natural and Applied Sciences*. 2010;4(3):236-54.
- [2] Choudhury S, Datta S, Talukdar AD, Choudhury MD. Phytochemistry of the Family Bignoniaceae-A review. *Assam University Journal of Science and Technology*. 2011;7(1):145-50.
- [3] Zuntini AR, Taylor CM, Lohmann LG. Deciphering the Neotropical *Bignonia binata* species complex (Bignoniaceae). *Phytotaxa*. 2015;219(1):69-77.
- [4] Zuntini AR, Taylor CM, Lohmann LG. Problematic specimens turn out to be two undescribed species of *Bignonia* (Bignoniaceae). *PhytoKeys*. 2015;(56):7-18.
- [5] Martin F, Hay AE, Corno L, Gupta MP, Hostettmann K. Iridoid glycosides from the stems of *Pithecoctenium crucigerum* (Bignoniaceae). *Phytochemistry*. 2007;68(9):1307-11.
- [6] da Rocha CQ, de-Faria FM, Marcourt L, Ebrahimi SN, Kitano BT, Ghilardi AF, et al. Gastroprotective effects of hydroethanolic root extract of *Arrabidaea brachypoda*: Evidences of cytoprotection and isolation of unusual glycosylated polyphenols. *Phytochemistry*. 2017;135:93-105.
- [7] Martin F, Hay AE, Cressend D, Reist M, Vivas L, Gupta MP, et al. Antioxidant C-glucosylxanthones from the leaves of *Arrabidaea patellifera*. *Journal of Natural Products*. 2008;71(11):1887-90.
- [8] Atawodi SEO, Olowoniyi OD. Pharmacological and therapeutic activities of *Kigelia africana* (Lam.) Benth. *Annual Research & Review in Biology*. 2015;5(1):1.
- [9] Saini S, Kaur H, Verma B, Singh S. *Kigelia africana* (Lam.) Benth. an overview. *Council of Scientific and Industrial Research*. 2009;8(2):190-197.
- [10] Houghton PJ, Osibogun IM. Flowering plants used against snakebite. *Journal of Ethnopharmacology*. 1993;39(1):1-29.
- [11] PioCorrea M. Dicionario das plantas Lcis do Brasil e das Exoticas cultivadas. *Zmprensa Nacional, Ministerio da Agricultura, IBDF, Rio de Janeiro, Brasil*. 1978;6:1926-54.
- [12] Aboutabl E, Hashem FA, Sleem A, Maamoon A. Flavonoids, anti-inflammatory activity and cytotoxicity of *Macfadyena unguis-cati* L. *African Journal of Traditional, Complementary and Alternative Medicines*. 2008;5(1):18-26.
- [13] Rice-Evans CA, Miller NJ, Paganga G. Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology and Medicine*. 1996;20(7):933-56.
- [14] Chen L, Chen D, Zheng Z, Liu S, Tong Q, Xiao J, et al. Cytotoxic and antioxidant activities of *Macfadyena unguis-cati* L. aerial parts and bioguided isolation of the antitumor active components. *Industrial Crops and Products*. 2017;107:531-8.
- [15] Castillo L, Díaz M, González-Coloma A, Rossini C. Differential activity against aphid settling of flavones obtained from *Clytostoma callistegioides* (Bignoniaceae). *Industrial Crops and Products*. 2013;44:618-21.
- [16] Bharti N, Singh S, Naqvi F, Azam A. Isolation and *in vitro* antiamoebic activity of iridoids isolated from *Kigelia pinnata*. *Arkivoc*. 2006;10(10):69-79.
- [17] Shah ZA, Khan MR. Peshawaraquinone a novel naphthoquinone and a new indanone from the stem of *Heterophragma adenophyllum* seem. *Records of Natural Products*. 2015;9(2):169.
- [18] Sidjui LS, Zeuko'o EM, Toghueo RMK, Noté OP, Mahiou-Leddet V, Herbette G, et al. Secondary metabolites from *Jacaranda mimosifolia* and *Kigelia africana* (Bignoniaceae) and their anticandidal activity. *Records of Natural Products*. 2014;8(3):307.
- [19] Ramirez G, Zamilpa A, Zavala M, Perez J, Morales D, Tortoriello J. Chrysoeriol and other polyphenols from *Tecoma stans* with lipase inhibitory activity. *Journal of Ethnopharmacology*. 2016;185:1-8.
- [20] Binutu OA, Adesogan KE, Okogun JI. Antibacterial and antifungal compounds from *Kigelia pinnata*. *Planta Medica*. 1996;62(04):352-3.
- [21] da Silveira JC, Gottlieb OR, de Oliveira GG. Zeyherol, a dillignol from *Zeyhera digitalis*. *Phytochemistry*. 1975;14(8):1829-30.
- [22] Zofou D, Tene M, Tane P, Titanji VP. Antimalarial drug interactions of compounds isolated from *Kigelia africana* (Bignoniaceae) and their synergism with artemether, against the multidrug-resistant W2mef *Plasmodium falciparum* strain. *Parasitology Research*. 2012;110(2):539-44.
- [23] Duarte D, Dolabela M, Salas C, Raslan D, Oliveiras A, Nenninger A, et al. Chemical Characterization and Biological Activity of *Macfadyena unguis-cati* (Bignoniaceae). *Journal of Pharmacy and Pharmacology*. 2000;52(3):347-52.
- [24] da Rocha CQ, Queiroz EF, Meira CS, Moreira DRM, Soares MBP, Marcourt L, et al. Dimeric flavonoids from *Arrabidaea brachypoda* and assessment of their anti-*Trypanosoma cruzi* activity. *Journal of Natural Products*. 2014;77(6):1345-50.
- [25] Zorn B, García-Piñeres AJ, Castro V, Murillo R, Mora G, Merfort I. 3-Desoxyanthocyanidins from *Arrabidaea chica*. *Phytochemistry*. 2001;56(8):831-5.
- [26] Sternitz FR, Arslanian RL, Castro O. Flavonoids from the leaf surface of *Godmania aesculifolia* (Bignoniaceae). *Biochemical Systematics and Ecology*. 1992;20(5):481.
- [27] Wollenweber E, Dörr M, Gomez LDP. Exudate flavonoids in *Godmania aesculifolia* (Bignoniaceae). *Biochemical Systematics and Ecology*. 1996;25(24):481-2.
- [28] Acerito T, Barbo FE, Negri G, Santos DYAC, Meda CI, Young MCM, et al. Foliar epicuticular wax of *Arrabidaea brachypoda*: flavonoids and antifungal activity. *Biochemical Systematics and Ecology*. 2002;30(7):677-83.
- [29] Pauletti PM, Bolzani VdS, Young MCM. Chemical constituents of *Arrabidaea samyoides* (Bignoniaceae). *Química Nova*. 2003;26(5):641-3.
- [30] Takemura OS, Inuma M, Tosa H, Miguel OG, Moreira EA, Nozawa Y. A flavone from leaves of *Arrabidaea chica* f. cuprea. *Phytochemistry*. 1995;38(5):1299-300.
- [31] Krebs HC. Inhaltsstoffe aus *Pseudocalymma elegans* (Vell.) Kuhlm. 1. Isolierung von Allantoin, Lamiid und Sorbarin/Ingredients from *Pseudocalymma elegans* (Vell.) Kuhlm. 1. Isolation of Allantoin, Lamiide, and Sorbarin. *Zeitschrift für Naturforschung B*. 1987;42(10):1361-4.
- [32] Jahangir M. Chemical and biological studies on some members of Asteraceae family and *Pseudocalymma elegans*, a native of Brazil: HEJ Research Institute of Chemistry, ICCBS, University of Karachi; 2001.
- [33] Leite JPV, Oliveira AB, Lombardi JA, S Filho JD, Chiari E. Trypanocidal activity of triterpenes from *Arrabidaea triplinervia* and derivatives. *Biological and Pharmaceutical Bulletin*. 2006;29(11):2307-9.
- [34] Govindachari TR, Patankar SJ, Viswanathan N. Isolation and structure of two new dihydroisocoumarins from *Kigelia pinnata*. *Phytochemistry*. 1971;10(7):1603-6.
- [35] Jassbi AR, Singh P, Jain S, Tahara S. Novel naphthoquinones from *Heterophragma adenophyllum*. *Helvetica Chimica Acta*. 2004;87(4):820-4.
- [36] Joshi KC, Singh P, Sharma MC. Quinones and other constituents of *Markhamia platycalyx* and *Bignonia unguis-cati*. *Journal of Natural Products*. 1985;48(1):145-.
- [37] Rocha CQd, Vilela FC, Santa-Cecilia FV, Cavalcante GP, Vilegas W, Giusti-Paiva A, et al. Oleanane-type triterpenoid: an anti-inflammatory compound of the roots *Arrabidaea brachypoda*. *Revista Brasileira de Farmacognosia*. 2015;25(3):228-32.
- [38] Atolani O, Olatunji GA, Fabiyi OA, Adeniji AJ, Ogbole OO. Phytochemicals from *Kigelia pinnata* leaves show antioxidant and anticancer potential on human cancer cell line. *Journal of Medicinal Food*. 2013;16(10):878-85.
- [39] Castillo L, Díaz M, González-Coloma A, González A, Alonso-Paz E, Bassagoda MJ, et al. *Clytostoma callistegioides* (Bignoniaceae) wax extract with activity on aphid settling. *Phytochemistry*. 2010;71(17-18):2052-7.
- [40] Tamayo-Castillo G, Vásquez V, Ríos MI, Rodríguez MV, Solano G, Zaccino S, et al. Isolation of major components from the roots of *Godmania aesculifolia* and determination of their antifungal activities. *Planta Medica*. 2013;79(18):1749-55.
- [41] Moideen S, Houghton PJ, Rock P, Croft S, Aboagye-Nyame F. Activity of extracts and naphthoquinones from *Kigelia pinnata* against *Trypanosoma brucei* brucei and *Trypanosoma brucei rhodesiense*. *Planta Medica*. 1999;65(06):536-40.
- [42] Joshi KC, Singh P, Taneja S, Cox PJ, Howie RA, Thomson RH. New terpenoid aldehydes from *Kigelia pinnata*: crystal structure of pinnatal. *Tetrahedron*. 1982;38(17):2703-8.
- [43] Jabeen B, Riaz N. Isolation and characterization of limonoids from *Kigelia africana*. *Zeitschrift für Naturforschung B*. 2013;68(9):1041-8.
- [44] Gouda YG, Abdel-baky AM, Darwish FM, Mohamed KM, Kasai R, Yamasaki K. Iridoids from *Kigelia pinnata* DC. fruits. *Phytochemistry*. 2003;63(8):887-92.

- [45] Houghton P, Akunyili D. Iridoids from *Kigelia pinnata* bark. *Fitoterapia-Milano*. 1993;64:473-.
- [46] Krebs HC. Inhaltsstoffe aus *Pseudocalymma elegans* (Vell.) Kuhlm. 2. Isolierung eines neuen Iridoidglucosids und von Durantosid II/Ingredients from *Pseudocalymma elegans* (Vell.) Kuhlm. 2. Isolation of a New Iridoid Glucoside and of Durantoside II. *Zeitschrift für Naturforschung B*. 1991;46(9):1258-60.
- [47] Bianco A, Massa M, Oguakwa JU, Passacantilli P. 5-Deoxystansioside, an iridoid glucoside from *Tecoma stans*. *Phytochemistry*. 1981;20(8):1871-2.
- [48] Lins AP, Felicio JDA. Monoterpene alkaloids from *Tecoma stans*. *Phytochemistry*. 1993;34(3):876-8.
- [49] Dickinson EM, Jones G. Pyridane alkaloids from *Tecoma stans*. *Tetrahedron*. 1969;25(7):1523-9.
- [50] Costantino L, Raimondi L, Pirisino R, Brunetti T, Pessotto P, Giannessi F, et al. Isolation and pharmacological activities of the *Tecoma stans* alkaloids. *Il Farmaco*. 2003;58(9):781-5.
- [51] Miranda N, Gerola AP, Novello CR, Ueda-Nakamura T, de Oliveira Silva S, Dias-Filho BP, et al. Pheophorbide a, a compound isolated from the leaves of *Arrabidaea chica*, induces photodynamic inactivation of *Trypanosoma cruzi*. *Photodiagnosis and Photodynamic Therapy*. 2017;19:256-65.
- [52] Kunapuli SP, Vaidyanathan CS. Indolic compounds in the leaves of *Tecoma stans*. *Phytochemistry*. 1984;23(8):1826-7.
- [53] de Oliveira AB, Isobe M, Braz-Filho R. Dipeptide from the roots of *Zeyhera digitalis*. *Journal of the Brazilian Chemical Society*. 1995;6(3):323-6.
- [54] Chivandi E, Cave E, Davidson BC, Erlwanger KH, Moyo D, Madziva MT. Suppression of Caco-2 and HEK-293 cell proliferation by *Kigelia africana*, *Mimusops zeyheri* and *Ximenia caffra* seed oils. *In Vivo*. 2012;26(1):99-105.
- [55] Verma PR, Deshpande SA, Rangari VD. Antinociceptive activity of aqueous extract of *Pachyptera hymenaea* (DC.) in mice. *Journal of Ethnopharmacology*. 2007;112(1):203-6.
- [56] Ufodike E, Omorogie E. Acute toxicity of water extracts of barks of *Balanites aegyptiaca* and *Kigelia africana* to *Oreochromis niloticus* (L.). *Aquaculture Research*. 1994;25(9):873-9.
- [57] Jothi ET, Harini CH, Reddy GN. Evaluation of letrozole induced polycystic ovary syndrome (pcos) using ethanol and ethyl acetate extracts of *Tecomaria capensis*. (family: Bignoniaceae). *World Journal of Pharmaceutical Research*. 2016;5(5):639-50.
- [58] Akhtar MS, Bashir S, Sial NT. Antimicrobial screening of *Heterophragma adenophyllum* extracts and effects of light irradiation. *Canadian Journal of Applied Sciences*. 2012;2(3):304.
- [59] Mafioletti L, da Silva Junior IF, Colodel EM, Flach A, de Oliveira Martins DT. Evaluation of the toxicity and antimicrobial activity of hydroethanolic extract of *Arrabidaea chica* (Humb. & Bonpl.) B. Verl. *Journal of Ethnopharmacology*. 2013;150(2):576-82.
- [60] Gaitán I, Paz AM, Zacchino SA, Tamayo G, Giménez A, Pinzón R, et al. Subcutaneous antifungal screening of Latin American plant extracts against *Sporothrix schenckii* and *Fonsecaea pedrosoi*. *Pharmaceutical Biology*. 2011;49(9):907-19.
- [61] Fournet A, Barrios AA, Munoz V. Leishmanicidal and trypanocidal activities of Bolivian medicinal plants. *Journal of Ethnopharmacology*. 1994;41(1-2):19-37.
- [62] da Rocha CQ, Vilela FC, Cavalcante GP, Santa-Cecília FV, Santos-e-Silva L, dos Santos MH, et al. Anti-inflammatory and antinociceptive effects of *Arrabidaea brachypoda* (DC.) Bureau roots. *Journal of Ethnopharmacology*. 2011;133(2):396-401.
- [63] Owolabi OJ, Omogbai EK. Analgesic and anti-inflammatory activities of the ethanolic stem bark extract of *Kigelia africana* (Bignoniaceae). *African Journal of Biotechnology*. 2007;6(5):582-5.
- [64] Michel AFRM, Melo MM, Campos PP, Oliveira MS, Oliveira FAS, Cassali GD, et al. Evaluation of anti-inflammatory, antiangiogenic and antiproliferative activities of *Arrabidaea chica* crude extracts. *Journal of Ethnopharmacology*. 2015;165:29-38.
- [65] Akah PA. Antidiarrheal activity of *Kigelia africana* in experimental animals. *Journal of Herbs, Spices & Medicinal Plants*. 1996;4(2):31-8.
- [66] Jackson SJ, Houghton PJ, Retsas S, Photiou A. *In vitro* cytotoxicity of norviburtinal and isopinnatal from *Kigelia pinnata* against cancer cell lines. *Planta Medica*. 2000;66(08):758-61.
- [67] Houghton PJ, Photiou A, Uddin S, Shah P, Browning M, Jackson SJ, et al. Activity of extracts of *Kigelia pinnata* against melanoma and renal carcinoma cell lines. *Planta Medica*. 1994;60(05):430-3.
- [68] Owolabi O, Amaechina F, Eledan A. Central nervous system stimulant effect of the ethanolic extract of *Kigelia africana*. *Journal of Medicinal Plants Research*. 2008;2(2):20-3.
- [69] Abioye A, Duru F, Noronha C, Okanlawon A. Aqueous extract of the bark of *Kigelia africana* reverses early testicular damage induced by methanol extract of *Carica papaya*. *Nigerian Journal of Health and Biomedical Sciences*. 2003;2(2):87-9.
- [70] Verma P, Deshpande S, Kamtham Y, Vaidya L. Hypolipidemic and antihyperlipidemic effects from an aqueous extract of *Pachyptera hymenaea* (DC.) leaves in rats. *Food chemistry*. 2012;132(3):1251-7.
- [71] Ittagi S, Merugumolu VK, Siddamsetty RS. Cardioprotective effect of hydroalcoholic extract of *Tecoma stans* flowers against isoproterenol induced myocardial infarction in rats. *Asian Pacific Journal of Tropical Disease*. 2014;4:S378-S84.
- [72] Granados-Echebogoyen C, Pérez-Pacheco R, Alexander-Aguilera A, Lagunez-Rivera L, Alonso-Hernández N, de Jesús Chairez-Martinez E. Effects of aqueous and ethanol extract of dried leaves of *Pseudocalymma alliaceum* (Bignonaceae) on haematological and biochemical parameters of wistar rats. *Asian Pacific Journal of Reproduction*. 2015;4(2):129-34.
- [73] Raju S, Kavimani S, Maheshwara Rao VU, Reddy KS, Kumar GV. Floral extract of *Tecoma stans*: A potent inhibitor of gentamicin-induced nephrotoxicity *in vivo*. *Asian Pacific Journal of Tropical Medicine*. 2011;4(9):680-5.
- [74] Ribeiro FM, Volpato H, Lazarin-Bidóia D, Desoti VC, de Souza RO, Fonseca MJV, et al. The extended production of UV-induced reactive oxygen species in L929 fibroblasts is attenuated by posttreatment with *Arrabidaea chica* through scavenging mechanisms. *Journal of Photochemistry and Photobiology B: Biology*. 2018;178:175-81.
- [75] Jorge MP, Madjarof C, Ruiz ALTG, Fernandes AT, Rodrigues RAF, de Oliveira Sousa IM, et al. Evaluation of wound healing properties of *Arrabidaea chica* Verlot extract. *Journal of Ethnopharmacology*. 2008;118(3):361-6.
- [76] Aro A, Freitas K, Foglio M, Carvalho J, Dolder H, Gomes L, et al. Effect of the *Arrabidaea chica* extract on collagen fiber organization during healing of partially transected tendon. *Life Sciences*. 2013;92(13):799-807.
- [77] Alonso-Castro AJ, Zapata-Bustos R, Romo-Yáñez J, Camarillo-Ledesma P, Gómez-Sánchez M, Salazar-Olivio LA. The antidiabetic plants *Tecoma stans* (L.) Juss. ex Kunth (Bignoniaceae) and *Teucrium cubense* Jacq (Lamiaceae) induce the incorporation of glucose in insulin-sensitive and insulin-resistant murine and human adipocytes. *Journal of Ethnopharmacology*. 2010;127(1):1-6.
- [78] Aguilar-Santamaría L, Ramírez G, Nicasio P, Alegría-Reyes C, Herrera-Arellano A. Antidiabetic activities of *Tecoma stans* (L.) Juss. ex Kunth. *Journal of Ethnopharmacology*. 2009;124(2):284-8.